

REMARKS

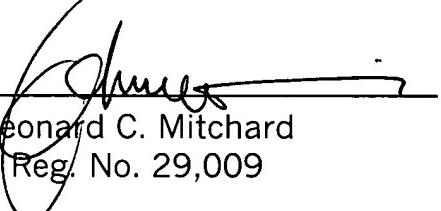
The specification and claims have been amended to conform with the attached sequence listing materials. No new matter is entered.

Attached hereto is a marked-up version of the changes made to the specification and claims by the current amendment. The attached page(s) is captioned "**Version With Markings To Show Changes Made.**"

Respectfully submitted,

NIXON & VANDERHYE P.C.

By: _____


Leonard C. Mitchard
Reg. No. 29,009

LCM:Iks
1100 North Glebe Road, 8th Floor
Arlington, VA 22201-4714
Telephone: (703) 816-4000
Facsimile: (703) 816-4100

VERSION WITH MARKINGS TO SHOW CHANGES MADE

IN THE SPECIFICATION

The paragraph beginning at page 8, line 7:

Figure 2. Best substrates identified by screening tyrosine library sub-sets 1 to 5 (SEQ ID NOS:4-7, respectively, in order of appearance) against ZAP-70 protein tyrosine kinase. The protein tyrosine kinase library described in Figure 1 was phosphorylated for 30 minutes at 30°C using the catalytic domain of human ZAP-70. Peptides were captured using streptavidin-coated 96 well plates and phosphotyrosine detected using anti-phosphotyrosine antibody, anti mouse IgG-HRP and tetramethylbenzidine (see experimental methods). Best substrates were identified as those which gave the highest amount of phosphate incorporation.

The paragraph beginning at Page 8, line 15:

Figure 3. Km Determination of Biotin-εAHA-DEEDYFE(Nle) [[SEQ ID NO. 3]] (SEQ ID NO:1). The catalytic domain of human ZAP-70 was used to phosphorylate varying concentrations of peptide for 10 minutes at 30°C in the presence of ^{33}P -γ-ATP. Peptide capture was performed using streptavidin filter plates, scintillation fluid added, and counting performed using a beta-counter (see experimental methods). Samples were assayed in triplicate.

The paragraph beginning at Page 9, line 16:

wherein each library subset is of formula (I)

(Xaa)_x Zaa (Xaa)_y (I) [[SEQ ID No. 1]]

The paragraph beginning at Page 10, line 5:

This invention can be applied for instance to a protein tyrosine kinase in order to exemplify the technology. It provides a rapid method of identifying discrete protein kinase substrate sequences which allows pharmacophore generation and design of active site inhibitors. This invention can also be used to directly identify protein kinase inhibitor molecules. General formula: (Xaa)_x Tyr (Xaa)_y(I) [[SEQ ID No. 2]]

The paragraph beginning at Page 11, line 25:

The results obtained from the library screen clearly demonstrated amino acid residues preferred by the protein kinase at each of the -4 to +4 sites (Figure 2). The 5 mer peptides overlapped to give information on amino acid preference at each of the binding positions -4 to +4. To confirm this a consensus peptide, Biotin-εAHA-DEEDYFE(Nle) [[SEQ ID No. 3]] (SEQ ID NO:1), representing the best -4 to +4 amino acids was made and tested as a substrate (Figure 3). This substrate gave a Km against ZAP-70 of 15.79 μM, which is better than the best ZAP-70 substrate described in the literature, a longer peptide of 14 amino acids with a tag of 3 arginines and a K_m of 29 μM (Wandenburg *et al*, 1996).

Page 13, line 18, amend to read:

General formula: (Xaa_x Ser (Xaa)_y [[SEQ ID No. 4]].

Page 19, line 6, amend to read:

5' CCGGGATCCGCCATGCCATGGACACGAGCGTGTAT 3' [[SEQ ID No. 5]] (SEQ ID NO:2)

Page 19, line 9, amend to read:

CTTCTGTGT 3' [(SEQ ID NO: 6)] (SEQ ID NO:3)

The paragraph beginning at Page 20, line 4:

Following homogenisation using a dounce pestle B, the cleared lysate was loaded onto a cobalt-sepharose column. After column washing with lysis buffer, elution was performed with an imidazole gradient and ZAP-70 fractions identified by protein kinase activity against the peptide substrate (SEQ ID NO:19) Lys-Lys-Lys-Lys-Ala-Asp-Glu-Glu-Asp-Tyr-Phe-Ile-Pro-Pro-Ala as described in Casnelli et al, 1991.

The paragraph beginning at page 20, line 28:

The best substrates were identified as those which gave the highest amount of phosphate incorporation. The library subsets were deconvoluted according to the teaching of W097/42216: this gives an immediate determination of the unique sequence of any phosphorylated motif without the need for further synthesis or sequencing. [(Figure 2 [SEQ ID Nos. 7,8,9,10])] (Figure 2 (SEQ ID NOS:4-7, respectively, in order of appearance)).

Page 21, line 25, amend to read:

Library Sub-Set 1 Asp-Glu-Glu-Asp-Tyr [[SEQ ID No. 11]] (SEQ ID NO:8)

Page 21, line 26, amend to read:

Library Sub-Set 2 Asp-Glu-Glu-Tyr-Asp [[SEQ ID No. 12]] (SEQ ID NO:9)

Page 22, line 1 amend to read:

Library Sub-Set 3 Asp-Glu-Tyr-Glu-Asp [[SEQ ID No. 13]] (SEQ ID NO:10)

Page 22, line 2 amend to read:

Library Sub-Set 4 Asp-Tyr-Glu-Glu-Val [[SEQ ID No. 14]] (SEQ ID NO:11)

Page 22, line 3 amend to read:

Library Sub-Set 5 Tyr-ser-Ile-Ile-Nle [[SEQ ID No. 15]] (SEQ ID

NO:12)

Page 22, line 12 amend to read:

Library Sub-Set 1 Asp-Glu-Glu-Glu-Tyr [[SEQ ID No. 16]] (SEQ ID NO:13)

Page 22, line 13 amend to read:

Library Sub-Set 2 Asp-Glu-Glu-Tyr-Phe [[SEQ ID No. 17]] (SEQ ID NO:14)

Page 22, line 14 amend to read:

Library Sub-Set 3 Asp-Glu-Tyr-His-Asn [[SEQ ID No. 18]] (SEQ ID NO:15)

Page 22, line 15 amend to read:

Library Sub-Set 4 Asp-Tyr-His-Leu-Phe [[SEQ ID No. 19]] (SEQ ID NO:16)

Page 22, line 16 amend to read:

Library Sub-Set 5 Tyr-Pro-Ile-Glu-Val [[SEQ ID No. 20]] (SEQ ID NO:17)

The paragraph beginning at Page 22, line 22:

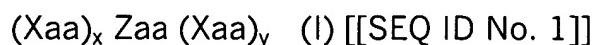
In this example the invention was used to map the active catalytic site of v-Abl, a protein kinase enzyme that catalyses the phosphorylation of a tyrosine residue. For this enzyme only the Library Sub-Set 4 (i.e. X-Tyr-X-X-X [according to SEQ ID No. 2]) was scanned with the enzyme. The active site substrate recognition substrate for this enzyme for this Sub-Set was Serine-tyrosine-phenylalanine-histamine-glutamine [[SEQ ID No. 21]] (SEQ ID NO:18).

IN THE CLAIMS

1. (Amended) A method for determining an amino acid sequence motif or a pentidomimetic sequence motif containing an active site capable of being bound by an enzyme which catalyses covalent modification of a substrate molecule, comprising:
 - a) contacting the enzyme with a library consisting of a number of oriented degenerate library subsets of molecules, each subset comprising unmodified degenerate motif sequences each having n residues and each having a modifiable residue at a different fixed non-degenerate position, under conditions which allow for modification of molecules which are a substrate for the enzyme;
 - b) allowing the enzyme to modify modifiable residues in library subsets containing molecules having an active substrate site for the enzyme;
 - c) deconvoluting the oriented degenerate library subsets of the library, *in situ* without separating modified from unmodified molecules, so as to reveal the

sequence of any motif which has been modified by covalent binding of the enzyme;

wherein each library subset is of formula (I)



wherein

Zaa is a non-degenerate modifiable natural or unnatural amino acid residue or peptidomimetic;

Xaa is any natural or unnatural amino acid residue or peptidomimetic;

x and y are each independently 0 or an integer;

(x + y) = n-1; and

n = an integer from 3 to 8, preferably 5.